α-HOMONOJIRIMYCIN [2,6-DIDEOXY-2,6-IMINO-D-GLYCERO-L-GULO-HEPTITOL] FROM <u>OMPHALEA DIANDRA L.</u>: ISOLATION AND GLUCOSIDASE INHIBITION

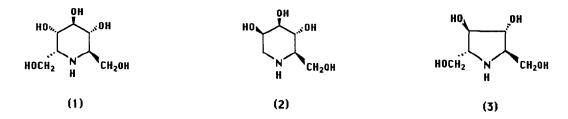
Geoffrey C. Kite,^a Linda E. Fellows,^a George W. J. Fleet,^b Paul S. Liu,^C Anthony M. Scofield^d and Neal G. Smith^e

^aJodrell Laboratory, Royal Botanic Gardens, Kew, Richmond, Surrey TW3 3DS, UK ^bDyson Perrins Laboratory, Oxford University, South Parks Road, Oxford, UK ^CMerrell Dow Research Institute, 2110, East Galbraith Road, Cincinnati, Ohio 45215, USA ^dDepartment of Biochemistry and Biological Sciences, University of London, Wye College, Ashford, Kent TN25 5AH ^eSmithsonian Tropical Research Institute, Box 2072, Balboa, Republic of Panama

The isolation of α -homonojirimycin [2,6-dideoxy-2,6-imino-D-glycero-L-gulo-heptitol] from <u>Omphalea diandra</u> is described; α -homonojirimycin is an inhibitor of several α -glucosidases.

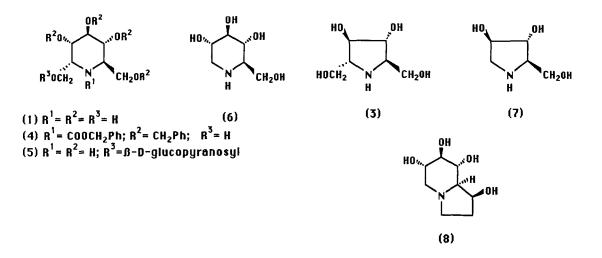
Alkaloidal glycosidase inhibitors, which are polyhydroxylated piperidines, pyrrolidines, octahydroindolizines and pyrrolizidines, have been found in a number of plants and microorganisms.^{1,2} In the angiosperms (flowering plants) these compounds have been reported in the families Leguminosae, Moraceae and Polygonaceae.³ Recently, the pyrrolidine glycosidase inhibitor DMDP [2,5-dideoxy-2,5-imino-D-mannitol] (3) was also found⁴ in a neotropical liana, <u>Omphalea diandra L.</u>, from the family Euphorbiaceae.

This paper describes the isolation from <u>Omphalea diandra L</u> of α -homonojirimycin [2,6-dideoxy-2,6-imino-D-glycero-L-gulo-heptitol] (1) and its ability to inhibit glycosidases; this is the first report of a naturally occurring azapyranose analogue of a heptose. <u>Omphalea diandra</u> has also been shown to contain deoxymannojirimycin [1,5-dideoxy-1,5-imino-D-mannitol] (2), an inhibitor of glycoprocessing mannosidase I and mammalian α -L-fucosidase, ⁵ previously isolated from the legume Lonchocarpus sericeus.⁶



Isolation. Leaves of Omphalea diandra L. were collected from plants growing wild in the Republic of Panama and oven dried at 60°C. Finely powdered leaf material was extracted into 70% aqueous methanol for 2 h at room temperature and the concentrated extract was passed down a column of Dowex 50 (H $^+$) ion exchange resin. Homonojirimycin was then eluted with 1 M pyridine, was freeze dried and the residue dissolved in water. Contaminating amino acids were removed by washing the solution through Amberlite CG-400 (OH-) ion exchange resin with water. Further purificaton by adsorption onto aluminum oxide 90 (Merck) and elution with acetone gave α -homonojirimycin, m.p. 206^o-207^oC, $[\alpha]^{20}$ (c, 0.54 in H_2^{0}) +88.2^o (589), +92.0^o (578), +103.9^o (546), +169.2^o (436) and +249.9^o (365); m/z (DCI NH₃): 194 (M+H⁺, 100%). ¹³C NMR (D₂O with dioxane as a internal standard): 54.77 (d), 57.06 (t), 57.57 (d), 67.72 (t), 72.26 (d), 72.77 (d) and 75.00 (d). This material had 1 H and 13 C NMR spectra which were superimposable on those of an authentic sample of synthetic α -homonojirimycin, m.p. 198⁰-199⁰C, $[\alpha]_{D}^{20}$ +79.1° (<u>c</u>, 2.03 in H₂0), prepared by hydrogenation of 2,6-dideoxy-2,6-[phenylmethoxycarbonyl]imino-1,3,4,5-tetra-O-benzyl-D-glycero-L-gulo-heptitol (4)⁷ or semi-synthetically from nojirimycin.⁸ Levels of homonojirimycin in O_{-} diandra leaves were estimated from GC analysis to be 0.3-0.4% of dry weight.9

After homonojirimycin had been eluted from the column of Dowex 50 (H^+) ion exchange resin by aqueous pyridine, further elution with 2 M aqueous ammonia gave a mixture¹⁰ containing deoxymannojirimycin (2) and DMDP (3).



Among naturally occurring azapyranose and azafuranose α -glucosidase inhibitors, α -homonojirimycin (1) is structurally related to deoxynojirimycin (6) in the same way that DMDP (3) is related to 1,4-dideoxy-1,4-imino-Darabinitol (7),¹¹ first isolated from <u>Angylocalyx</u>.¹² The B-D-glucopyranosyl derivative (5) of homonojirimycin was designed^{7,8} as a synthetic transition state inhibitor of α -glucohydrolases and has been shown to be an effective means of reducing the hyperglycemic response to an oral sucrose or starch load and may therefore have potential as a drug for the treatment of diabetes mellitus.¹³ Also, several α -glucosidase inhibitors, including castanospermine (8), isolated from the seeds of the Australian legume <u>Castanospermum australe</u>,¹⁴ inhibit human immunodeficiency virus syncytium formation and virus replication;¹⁵ such compounds may have potential as antiretroviral agents.¹⁶

 α -Homonojirimycin was tested as a possible inhibitor of glycosidase activity in homogenates of mammalian (mouse) and insect (<u>Spodoptera littoralis</u>, fifth and sixth instar larvae) gut, using both synthetic (<u>p</u>-nitrophenyl glycopyranosides) and natural substrates. The assay methods employed have been described elsewhere.^{2,17}

TABLE. Concentration of α -homonojirimycin (M) giving 50% inhibition of glycosidase activity [N.I. = less than 50% inhibition at 3.3 x 10⁻⁴M]

Substrate		genate from: podoptera littoralis
p-nitrophenyl α-D-glucopyranoside	2.2×10^{-7}	1.3×10^{-5}
p-nitrophenyl ß-D-glucopyranoside	1.4×10^{-4}	N. I.
<u>p</u> -nitrophenyl α -D-galactopyranoside	5.3×10^{-5}	N. I.
maltose	7.2×10^{-7}	1.3×10^{-4}
sucrose	8.1×10^{-8}	2.2×10^{-4}
lactose	5.5×10^{-5}	1.9×10^{-4}
trehalose	1.8×10^{-5}	3.7×10^{-5}

Thus, α -homonojirimycin (1) was found to be a potent inhibitor (Table) of digestive α -glucosidase activity in mouse but, with the exception of trehalase, to be relatively weak against comparable activity in <u>Spodoptera</u>. In this respect, α -homonojirimycin resembles deoxynojirimycin (6);^{1,17} its potential as an antiviral and anti-diabetic agent is under investigation.

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